

AEROBIC BACTERIOLOGICAL PROFILE OF SKIN AND SOFT TISSUE INFECTIONS (SSTI'S) AND IT'S ANTIMICROBIAL SUSCEPTIBILITY PATTERN AT M. B. GOVT. HOSPITAL IN UDAIPUR, RAJASTHAN

Anshu Sharma¹, Sandeep Gupta^{2*}

¹ Professor, Department of Microbiology, RNT Medical College, ² Resident, Department of Microbiology, RNT Medical College, Udaipur, Rajasthan

*Email id of corresponding author- drsandip80@gmail.com

Received: 15/01/2016

Revised: 26/02/2016

Accepted: 26/04/2016

Abstract:- Introduction: Skin and soft tissue infections (SSTI's) are a common type of infection. The overall incidence of wound sepsis in India is from 10-33%. A predictable bacterial profile in the wound infections is very important for clinicians, who intend to start empirical treatment for patients, while laboratory culture reports are awaited. **Objectives:** To identify the common aerobic bacterial isolates and their antimicrobial susceptibility pattern along with methicillin resistance in Staphylococcus aureus and CoNS, production of extended spectrum beta lactamases in Escherichia coli and Klebsiella spp and Biofilm production in Staphylococcus aureus and CoNS. **Material and Methods:** A total of randomly 207 pus samples received for aerobic bacteriological culture and sensitivity in the microbiology department of R.N.T. Medical College Udaipur, from various departments (OPD/IPD) of M.B.G. Hospital, Udaipur. This prospective study was done the period of six month (Dec.2014 to June 2015). The samples were processed in the laboratory by standard techniques. **Results:** Out of 207 samples 178(86%) were positive for aerobic bacterial culture while 29 (14%) samples had no growth. Among the 178 culture positive pus samples, 151(84.83%) yielded pure bacterial isolates and 27(15.17%) yielded mixed infection. In the present study, Staphylococcus aureus 77 (37.5%) was the commonest organism isolated. Out of 77 Staphylococcus aureus isolated, 31 (40.25%) were MRSA and ESBL producers among Escherichia coli and Klebsiella species were 30 (85.71%) and 4(50%) respectively. Out of 103 Staphylococcus aureus (77) and CoNS (26) isolates, 33 (32.03%) were biofilm producer. **Conclusion:** The commonest isolates of skin and soft tissue infections are Staphylococcus Aureus (37.5%) followed by Escherichia Coli (17%), CoNS (13%) and Pseudomonas spp (12%) and there is a high level of resistance against commonly used antimicrobials due to methicillin resistance, ESBL and Biofilm production. Staphylococcus aureus were 100% sensitive to vancomycin and Escherichia coli were 80% sensitive to amikacin so these antimicrobials can be included in empirical treatment of skin and soft tissue infections. **Keywords:** Antimicrobial Susceptibility, Biofilm, ESBL, MRSA, Staphylococcus aureus, skin and soft tissue infections (SSTI'S).

INTRODUCTION:

Skin and soft tissue infections (SSTI's) are a common type of infection. Common example SSTIs includes cellulitis, abscesses,

impetigo, folliculitis, furuncle, carbuncle, necrotizing fasciitis, diabetic foot ulcer and surgical site infections. The development of

wound infection depends on the integrity and prospective function of skin.(1) The chances of infection depends on patient conditions such as the state of nutrition and existing diabetes mellitus like medical conditions and pre, intra and postoperative care if the patient has undergone surgery. Thus it is difficult to predict which wound will become infected.(2) In India incidence of wound sepsis is 10-33%.(3) Wound infections can be caused by different groups of microorganisms, most commonly isolated aerobic microorganisms includes Staphylococcus aureus, Coagulase negative staphylococcus, Enterococci, Escherichia coli, Pseudomonas aeruginosa, Klebsiella pneumoniae, Enterobacter, Proteus mirabilis, other streptococci, Candida and Acinetobacter.(4) Coagulase positive Staphylococcus aureus has been found to be more dominant organism in pus.(5) Staphylococci are ubiquitous and most common cause of localized suppurative lesion in human beings.

Wound infection is one of the most common hospital acquired infections and important cause of morbidity and accounts for 70-80 %.(6) Developments of such infections represent delayed healing, cause anxiety and discomfort for patient. The importance of wound infections in both economic and human terms, should not be underestimated.(7) In a study on an average, patients with wound infections stays about 6-10days more in hospital than if the wound had heal without infections.(8) This additional stay doubles the hospital cost.

Relative resistance to antibiotics relatively more virulent strains and capacity to adapt quickly to changing environment make the pathogens acquired in hospitals a matter of concern.(9) Effective treatment of wound infections depends upon proper understanding of

causative pathogen, pathophysiology of the infectious process and pharmacology of the therapeutic agents. The inadvertent use of antibiotics leads to emergence of drug resistant pathogens, which in turns acts as a great challenge to the health services.

A predictable bacterial profile in the wound infections is very important for clinicians who intend to start empirical treatment for patients, while laboratory culture reports are awaited. The purpose of this study is to show the spectrum of aerobic bacterial profile and its sensitivity pattern from skin and soft tissues infections.

MATERIAL AND METHOD:-

A total of randomly 207 pus samples received for aerobic bacteriological culture and sensitivity in the bacteriology section of microbiology department of R.N.T. Medical College Udaipur, from various departments (OPD/IPD) of M.B.G. Hospital, Udaipur. This study was done the period of six month (Dec.2014 to June 2015). The samples were transported immediately and processed in the laboratory as per standard protocol.(10) Gram staining was done and the samples were inoculated into blood agar, MacConkey agar and Glucose Broth by standard techniques. The plates were incubated at 37°C for 24-48 hours and growth was observed. On correlating the gram stain and culture report, further identification were done with biochemical tests(10) such as catalase, coagulase, oxidase, indole, methyl red, voges-proskauer, citrate, urease, triple sugar iron, mannitol salt agar, motility by hanging drop method, sugar fermentation test and Amino acid decarboxylase and Arginine dihydrolase test. Kirby-Bauer disc diffusion assay on Mueller

Hinton agar were carried out to determine the antimicrobial susceptibility profiles as per CLSI guidelines.(11,12) Separate set of antibiotic were used for gram positive organisms and gram negative organisms. Detection of Methicillin Resistant Staphylococcus Aureus (MRSA) was done with Oxacillin (1µg) disc by disc diffusion test.(13) Extended spectrum β- lactamases (ESBL) production in *E. coli* and *Klebsiella* spp was detected by phenotypic confirmatory test(14) with the help of Ceftazidime (30 µg) discs alone and in combination with Clavulanic acid (Ceftazidime + Clavulanic Acid, 30/10 µg) disc. The control strains used were *Escherichia coli* ATCC 25922 as a non-ESBL producing organism and *Klebsiella pneumoniae* ATCC 700603 as an ESBL producing organism. Biofilm production was determined by using two methods, Congo red agar method(15) and Christensen's tube method.(16) Biofilm producing *Staphylococcus epidermidis* ATCC 35984 used as positive control and negative control was un-inoculated plate and tube respectively.

RESULTS :-

The period of study is from December 2014 to June 2015 with total of 207 pus samples. Out of 207 samples 178(86%) were positive for aerobic bacterial culture while 29 (14%) samples had no growth. Of the positive culture, 129 (72.48%) patients were males and 49 (27.52%) were females yielding a ratio of 2.63 (Graph 1). Among the 178 culture positive pus samples, 151(84.83%) yielded pure bacterial (mono-microbial) isolates and 27(15.17%) yielded mixed infection (two organisms- polymicrobial); so a total number of 205 organisms were isolated out of 207 pus samples (Graph 2). The

department wise distribution of pus samples revealed that surgery dept. was the highest contributor (40%), followed by Skin & V.D.(32%), ENT (16%), Obs & Gynae (5%) (Table 1).

In the present study among the 205 isolates, *Staphylococcus aureus* was the commonest organism isolated. Table shows that *Staphylococcus aureus* was isolated in 77 samples accounting for 37.5% in the total isolated organism. The second commonest organism was *E. coli* 35 (17%) followed by Coagulase negative staphylococcus 26 (13%), *Pseudomonas* spp. 24 (12%), *Citrobacter* spp 13 (6%), *Klebsiella* spp 8 (4%), *Enterococcus faecalis* 6(3%), *Streptococcus* spp 3 (2%), *Proteus mirabilis* 2 (1%), *Acinetobacter* spp 3 (1%), *Enterobacter* spp 2 (1%), GPB 4 (2%) and *Candida albicans* was observed in 1 cases (Table 2).

The Antibiogram of Gram Positive Cocci (Table 3) revealed that the Vancomycin (99%) was the most susceptible drug followed by Gentamicin (85%), Amikacin (82%), Doxycycline (78%), Tetracycline (78%) and Oxacillin (58%). Gram Negative Bacilli are most susceptible to Amikacin(76%) followed by Ceftazidime + Clavulanic acid (74%), Gentamicin (63%), Ciprofloxacin (47%) and Carbenicilline (39%) (Table 4).

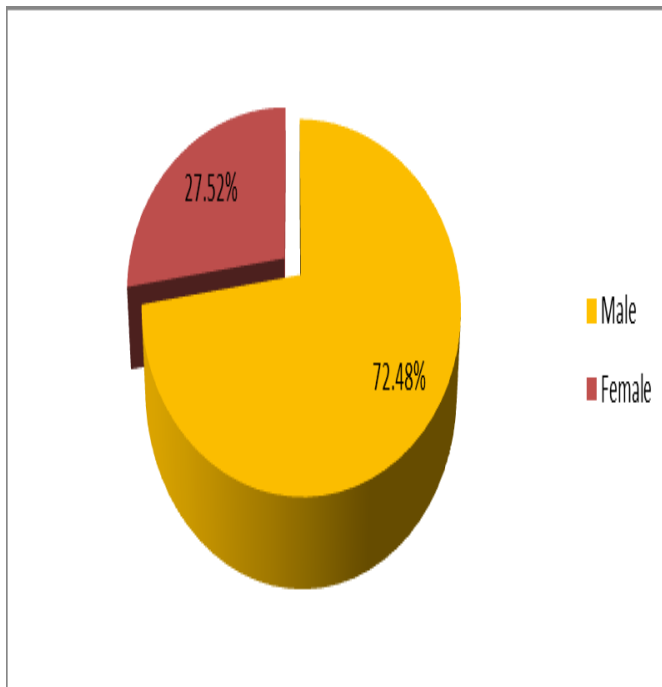
MRSA was detected with the help of Oxacillin discs (1µg) and Out of 77 *Staphylococcus aureus* isolated, 31 (40.25%) were MRSA and 46 (59.75%) were MSSA and among CoNS (26), Methicillin resistance CoNS were 13(50%) (Graph 3, 4).

ESBL producers among *Escherichia coli* and *Klebsiella* species was detected with Ceftazidime (30 µg) and Ceftazidime + Clavulanic acid (30/10 µg) disc and found to be

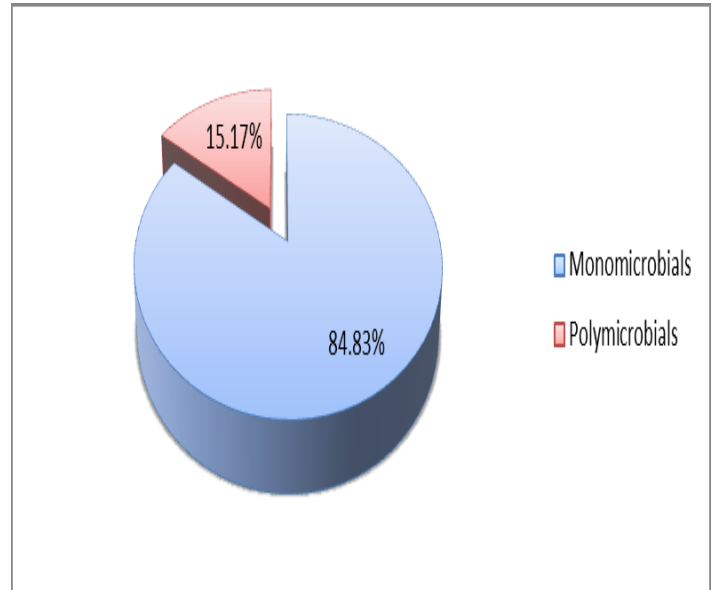
30 (85.71%) and 4(50%) respectively (**Graph 5, 6**).

Out of 103 *S. aureus* (77) and CoNS (26) isolates, 33 (32.03%) were biofilm producer and 70 (67.97%) were non-biofilm producer (**Table 5**). Biofilm production was more in MRSA (54.83%) than MR CoNS (38.46%) in present study. Biofilm production was more in Methicillin resistant strains than Methicillin sensitive strains in both *Staphylococcus aureus* and CoNS (**Table 6**).

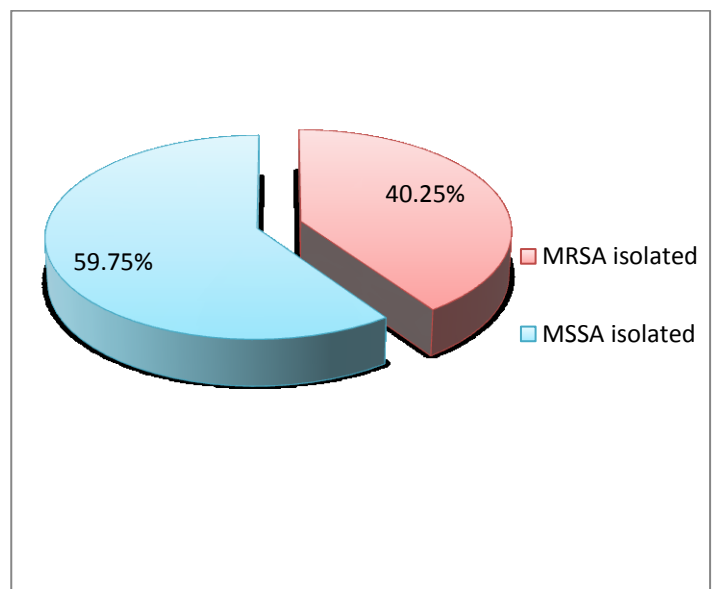
Graph 1: Showing Sex wise distribution



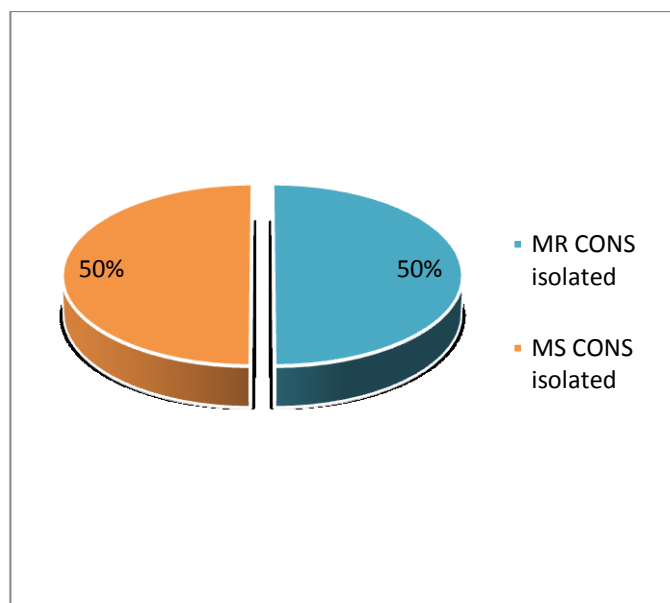
Graph 2: Percentage of Pure and Mixed Cultures



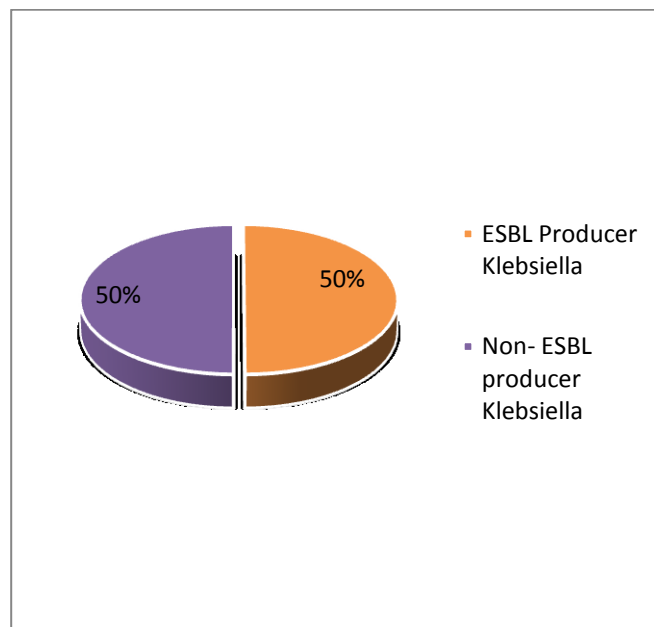
Graph 3: Staphylococcus aureus: MRSA Producer



**Graph 4: Coagulase Negative
Staphylococcus (CoNS) isolates:
Methicillin resistant**



**Graph 6: ESBL producing and Non-ESBL
producing Klebsiella spp**



**Graph 5: ESBL Producer and non-ESBL
Producer in E. coli**

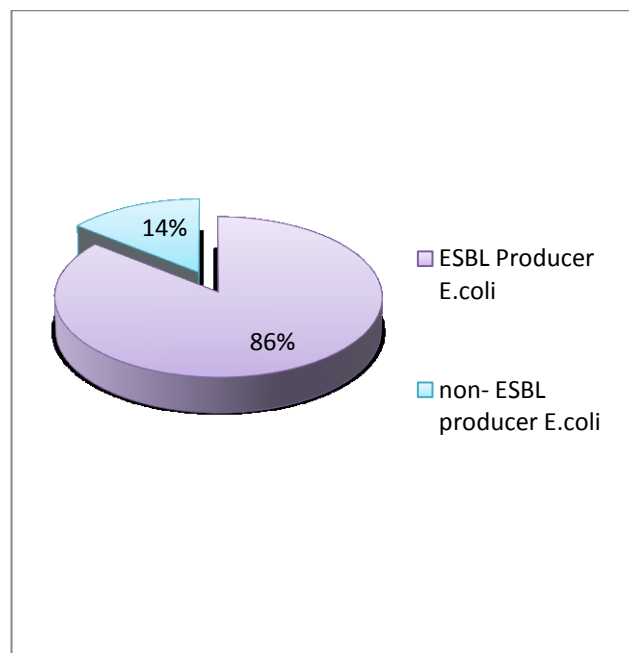


Table 1: Department wise distribution

Name of Department	No. of patients	Percentage
General Surgery	82	40%
Skin & V.D.	66	32%
ENT	33	16%
Obs. & Gynae.	11	5%
Radiotherapy	5	2%
Neurosurgery	4	2%
Orthopedics	3	1%
Paediatric surgery	2	1%
Medicine	1	1%
Total	207	100%

Table 2: Percentage of the total organisms (Mono-microbial + Poly-microbial) isolates in the present study

S. No.	Organisms	No. of organisms (n=205)	Percentage (%) (n=205)
1	Staphylococcus aureus	77	37.5%
2	Escherichia coli	35	17%
3	CoNS	26	13%
4	Pseudomonas spp	24	12%
5	Citrobacter spp	13	6%
6	Klebsiella spp	8	4%
7	Enterococcus spp	6	3%
8	Streptococcus spp	4	2%
19	Acinetobacter spp	3	1%
10	Proteus spp	2	1%
11	Enterobacter spp	2	1%
12	Candida albicans	1	0.5%
13	GPB	4	2%
	Total	205	100%

Table 3: Antibiotic susceptibility pattern of different gram positive cocci (n=113)

S.NO	Antibiotic	Staphylococcus aureus n=77	CoNS n=26	Enterococcus spp n=6	Streptococcus spp n=4
1	AK	88%	73%	50%	75%
2	GEN	86%	88%	50%	100%
3	CN	49%	50%	33%	100%
4	CTX	56%	50%	50%	100%
5	CAZ	39%	38%	17%	75%
6	CTR	51%	50%	50%	100%
7	ERY	31%	38%	17%	100%
8	AMX	13%	23%	0%	100%
9	AMC	27%	35%	50%	100%
10	CIP	51%	50%	50%	100%
11	COT	25%	38%	0%	25%
12	DO	81%	77%	50%	75%
13	TE	81%	77%	50%	75%
14	OX	60%	50%	50%	75%
15	VAN	100%	100%	83%	100%

Table 4: Antibiotic susceptibility pattern of different gram negative bacilli (n=87)

S.NO	Antibiotic	E.coli n=35	Pseudomonas spp n=24	Citrobacter spp n=13	Klebsiella spp n=8	Acinetobacter spp n=3	Enterobacter spp n=2	Proteus spp n=2
1	AK	66%	75%	69%	75%	67%	50%	100%
2	GEN	57%	63%	46%	75%	33%	50%	100%
3	CN	3%	0%	23%	25%	0%	0%	0%
4	CTX	3%	63%	23%	50%	33%	50%	0%
5	CAZ	3%	67%	23%	50%	33%	50%	0%
6	CTR	3%	54%	23%	50%	33%	50%	0%
7	AMX	3%	4%	15%	0%	0%	0%	0%
8	AMC	3%	4%	15%	25%	0%	0%	100%
9	CIP	9%	79%	54%	63%	33%	50%	100%
10	COT	17%	13%	46%	63%	0%	50%	100%
11	DO	23%	4%	46%	88%	67%	50%	50%
12	NA	0%	0%	15%	38%	0%	0%	0%
13	TE	23%	0%	46%	88%	67%	50%	50%
14	CB	6%	88%	23%	50%	33%	50%	50%
15	CAC	43%	92%	85%	88%	33%	100%	100%

Table 5: Shows distribution of biofilm producing strains (Staphylococcus aureus & CoNS)

Total isolates=103			
Biofilm producers=33		Non biofilm producers=70	
Staphylococcus aureus=27	CoNS=6	Staphylococcus aureus=50	CoNS=20

Table 6: Shows relation of methicillin resistance in relation to biofilm production

Biofilm production	MRSA	MSSA	MR CoNS	MS CoNS
Biofilm producers	17	10	5	1
Non-biofilm producers	14	36	8	12
% Biofilm producers	54.83	21.73	38.46	7.7
% Non-biofilm producers	45.17	78.27	61.54	92.3

DISCUSSION:

Infection of wound is one of the common cause to increase hospital stay and treatment cost. Emerging drug resistant organisms further increases the threat. The identification of common causative bacteria of wound infections

with their antimicrobial sensitivity pattern will be helpful to the clinician in choosing an empirical antibiotic therapy. In the present study an attempt was made to know the identification of various pus isolates with their antibiotic susceptibility testing. In our study, the wound infection was more common in male than female (2.63:1). N.

Sowmya and S. Savitha et al.(17) (2014) also observed males to be commonly infected than females (2:1). In the present study incidence of wound isolates cases was more common in General surgery department (40%) followed by Skin & V.D. department (32%), ENT (16%), Obs. & Gynae. (5%), Radiotherapy (2%) and Neurosurgery (2%). My study is correlated with other workers like V.M.V.S.V Raghav Rao et al.(18) (2014) have found the department wise distribution of pus samples that revealed that surgery dept. was the highest contributors (35.29%), followed by Orthopedics (29.42%), Gynae & Obs. (11.76%), Medicine (9.80%), Skin (7.85%) and ENT (5.88%) departments.

Majority of our results are mono-microbial (85%) and *Staphylococcus aureus* was found to be the most common pathogen in our study (37.5%), similar reports was also observed by N. Sowmya and S. Savitha et al.(17) (2014), A.Ananth and S.Rajan(19) (2014), A.R.Kumar(20) (2013). The second common pathogen in our study was *E.coli* (17%). D.V.M.V.S.V et al.(18) (2014) and S.Mohanty and A.Kapil et al.(21) (2004), also reported *Escherichia coli* to be the second most commonly occurring pathogens in wound infections. Gram positive organisms obtained in our study were 99% sensitive to vancomycin. In our study, MRSA (Oxacillin resistant) accounts for 40.25% which is in comparison with other workers like N. Lakshmi et al.(22) (2015) 39.1%, INSAR (2008) 36% in pus, INSAR (2009) 40% and Rajeshwar et al.(23) (2014) 32.70%. So overall we see that the prevalence of MRSA is similar or on a higher level in our region as compared to other studies ranging from 27-40%.

In present study, *Escherichia coli* is 80% sensitive to amikacin, 69% sensitive to gentamicin, 54% sensitive to ceftazidime +

clavulanic acid and 34% sensitive to tetracycline and doxycycline followed by cotrimoxazole 29%.

In our study ESBL production was higher in *E. coli* in comparison to *Klebsiella* spp. Out of the total 35 *E. coli* isolates ESBL production was seen in 30 (85.71%) *Escherichia coli* isolates. While out of 8 *Klebsiella* spp. isolates ESBL production was seen in 4 (50%) isolates. ESBL producers *Escherichia coli* accounts for 34.48% among GNB which is in comparison with other workers like Rajeshwar Rao et al.(23) (2014) which showed Extended Spectrum Beta Lactamase were found in variety of Enterobacteriaceae members (30.9%). B Fouzia et al.(24) (2013) demonstrate Enterobacteriaceae family showed 32% ESBL (Extended spectrum beta lactamase) producer.

In our study, we isolated 33(32%) biofilm producers out of 103 GPC (*Staphylococcus aureus* and CoNS) isolates. In which 27(81.81%) were *Staphylococcus aureus* and 6 (18.19%) were CoNS. Biofilm production was observed in 54.83%(17/31) MRSA and 21.73%(10/46) MSSA while 38.46%(5/13) MRCoNS and 7.69%(1/13) MSCoNS were found to be biofilm producers. A study by Charankaur et al.(25) concludes 78.8% MRSA as biofilm producer in various clinical isolates in Pune. Maximum biofilm production was seen in Pus samples followed by Urine. Aggrawal et al.(26) reported 79% biofilm producing *Staphylococcus aureus* and 43% biofilm producing CoNS from blood samples in Lukhnow. 82% of biofilm producing *S. aureus* and 71.4% biofilm positive CoNS were methicillin resistant which is higher in no. as compared to our study.

Biofilm producer *S. aureus* showed maximum susceptibility to Vancomycin (100%), doxycycline, tetracycline, amikacin (81.48%)

each and gentamicin (77.77%) followed by ciprofloxacin (44.44%), cefotaxime and oxacillin (37.03%). On the contrary, non-biofilm producing *S. aureus* were comparatively much more sensitive to these antimicrobials.

CONCLUSION: -

This study revealed the presence of wound infection causing bacteria, those are capable of causing various human illness. The bacterial isolate screened in various skin and soft tissue infections collected from various wards. A total of randomly 207 samples received in the bacteriology section of microbiology department of R.N.T. Medical College, Udaipur, from various departments (OPD/IPD) of M.B.G. Hospital, Udaipur.

The commonest isolates of Wound infection are *Staphylococcus Aureus* (37.5%) followed by *Escherichia Coli* (17%), CoNS (13%) and *Pseudomonas spp* (12%) and there is a high level of resistance against commonly used antimicrobials.

Among the *Staphylococcus aureus*, MRSA accounts for 40.25% and among CoNS Methicillin resistance accounts for 50%. Methicillin resistant *Staphylococcus aureus* (MRSA) is now endemic in India. The incidence of MRSA varies from 25 per cent in western part of India and 2 to 50 per cent in South India. Community acquired MRSA (CA-MRSA) has been increasingly reported from India.(27) The prolonged hospital stay, indiscriminate use of antibiotics, lack of awareness, receipt of antibiotics before coming to the hospital etc. are the possible predisposing factors of MRSA emergence.

Among Gram negative bacilli, ESBL producer *Escherichia coli* accounts for 34.48% and ESBL producer *Klebsiella spp* accounts for 4.59%. Among *Escherichia coli*, ESBL producer

E. coli accounts for 85.71% and among *Klebsiella spp*, ESBL producer *Klebsiella spp* accounts for 50%.

Out of 103 *Staphylococcus aureus* (77) and CoNS (26) isolates, 33(32.03%) were biofilm producer and 70 (67.97%) were non-biofilm producer. Biofilm production was more in MRSA (54.83%) than MRCoNS (38.46%) in present study. Biofilm production was more in Methicillin resistant strains than Methicillin sensitive strains in both *Staphylococcus aureus* and CoNS.

In our study biofilm production was lower than other studies, so it is right time to take action like strict implementation of good infection control programs against spreading of these more resistant biofilm producing isolates.

CONCLUSION

Hence, Knowledge of the most common causative agents of infection and their antimicrobial susceptibility pattern is very essential for the judicious administration of empirical treatment before the culture results are available. Antimicrobial susceptibility of microorganisms varies from time to time and from place to place. Hence regular monitoring of bacterial susceptibility to antibiotics in skin and soft tissues infections is essential for appropriate therapy. Good infection control programs are to be maintained and avoid indiscriminate use of broad spectrum antibiotics to curtail emergence of MRSA, ESBL and Biofilm producers. Antibiogram should be prepared regularly and made readily available to the clinicians to guide them in therapy. There is a need for a central database in India where various laboratories can upload their antibiogram regularly and this data

can be very useful in formulating guidelines for treatment of various infectious diseases.

Funding: No funding sources

Conflict of interest: None declared

Ethical approval: The study was approved by the institutional ethics committee

REFERENCES: -

1. Calvin M. Cutaneous wound repair. *Wounds*. 1998; 10:12-14
2. Karia JB, Gadekar HB, Lakhani SJ. Study of bacterial profile of pus culture in Dhiraj general Hospital. www.themedicalacademy.in
3. Basu S, Ramchuran Panray T, Bali Singh T, Gulati AK, Shukla VK. A prospective, descriptive study to identify the microbiological profile of chronic wounds in outpatients. *Ostomy Wound Manage*. 2009 ; 55: 14-20.
4. Tayfour MA , Al-Ghamdi SM and Al-ahamdi AS. Surgical wound infections in King Fahed Hospital at Al-baha Saudi Med.J. 2005;26(8):1305-07
5. Chopra, A., Puri, R., Mittal, R. R. and Kanta, S. 1994. A clinical and bacteriological study of pyodermas. *Indian J. Dermatology, Venerology and Leprology*. 60: 200-202.
6. Collier M. Recognition and management of wound infections *Wounds*. available from URL:<http://www.worldwidewounds.com>.
7. Plowman R. The Socio economic burden of hospital acquired infection. *Euro saweill*. 2005; 5(4):49-50.
8. Henzelmann M, Scott M, Lam T. Factors predisposing to bacterial invasion and infection. *Am J Surg* 2002;183(2):179-90
9. Plummer D. Surgical Wound infections as a performance indicator: agreement of common definitions of wound infections in 4773 patients. *BMJ*. 2004;329:720-22
10. Collee J.G., Fraser A.G. Mackaie & MacCarteny, *Practical Medical Microbiology*. 14th edition
11. Elmer .Koneman, Stephan Dallen, W.M Janda, PC Schreckenberger. *color atlas and textbook of diagnostic microbiology*, 5th edition San Francisco Lippincott, 1997; 539-576
12. Clinical and laboratory standard institute. Performance standards for antimicrobial susceptibility testing.M100-S17, 2007; 27(1).
13. Choudhury R, Panda S, Singh DV. Emergence and dissemination of antibiotic resistance: A global problem. *Ind. Med. Microbiol*. 2012; 30(4): 384-90.
14. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing. Twentieth informational supplement ed. CLSI document M100-S20. Wayne, PA: CLSI; 2010.
15. Freeman D J, Falkiner F R, Keane C T. New method for detecting slime production by coagulase negative staphylococci. *J Clin Pathol* 1989; 42:872-874.
16. Gordon D. Christensen, W. Andrew Simpson, Alan L. Bisno and Edwin H. Beachey. Adherence of Slime-Producing Strains of *Staphylococcus epidermidis* to Smooth Surfaces. *Infect. Immun*. 1982, 37(1):318.
17. Sowmya N, Savitha S, Mallure S, Mohankrishanan K, Sumathi G and Arumugan P . A two year study of spectrum of bacterial isolates from wound infections

- by aerobic culture and their antibiotic pattern in a tertiary care center. Int J Curr. Microbiol. and App. Sci. IISN: 2319-7706 vol. 3 November 8 (2014):292-295
18. Raghav Rao D.V.M.V.S.V, Basu R, Debika Roy Biswas, Aerobic Bacterial Profile and Antimicrobial Susceptibility Pattern of Pus Isolates in a South Indian Tertiary Care Hospital. Journal of Den.and Med. Sci.2014; 13: 59-62.
 19. Ananth A and Rajan S. Isolation and Screening of Pathogenic Bacteria from Wound Infections. Int. J. of Curr. Pharmaceutical Research.2014; 6(3): 15-17.
 20. Kumar A R. Antimicrobial Sensitivity Pattern of *Staphylococcus aureus* isolated from Pus from tertiary Care Hospital, Surendranagar, Gujarat and issues related to the rational selection of antimicrobials. Scholars Journal of App. Med. Sci. 2013; 1(5):600-605.
 21. Mohanty S, Kapil A, Dhawan B, Das BK. Bacteriological and antimicrobial susceptibility profile of soft tissue infections from northern India. Ind. J. Med. Sci. Jan 2004; 58(1): 10-15.
 22. Lakshmi N, Koripella R, Manem J, Balamurali Krishna P. Bacteriological Profile and Antibigram of Burn Wound Infections in a tertiary care hospital. J. Dent. and Med. Sci. (IOSR-JDMS) 2015;14(10): 01-04.
 23. Rajeshwar Rao S, Jaya Lakshmi L, Pavani S, Kawle V and Jaya Prakash S. Bacteriological Profile, Antibigram of Burn wound Isolates and Detection of MRSA and ESBL Production at Tertiary Care Hospital, Hyderabad. World J. Pharma. and Pharmaceuticals Sci. 2014; 3:1691-1698.
 24. Fouzia B, Damle A S and G Maher. Changing patterns of burn infections. IOSR J. Den. and Med. Sci.2013; 5:11-14.
 25. Dardi Charan Kaur, Khare A.S. Biofilm formation and antibiotic susceptibility pattern in MRSA strains in a tertiary care rural hospital. Ind. J. of Basic and App. Med. Res.; December 2013; 3(1): 37-44.
 26. Agarwal A, Jain A . Association between drug resistance & production of biofilm in staphylococci. Ind. J. Med. Res. April 2012; 135:562-564.
 27. Anand KB, Agrawal P, Kumar S, Kapila K. Comparison of cefoxitin disc diffusion test, oxacillin screen agar, and PCR for *mecA* gene for detection of MRSA. Ind. J. Med. Microbiol. 2009; 27(1): 27-9.